



Geranylgeraniol Supplementation Mitigates Soleus Muscle Atrophy via Changes in Mitochondrial Quality Control in Diabetic Rats



Nigel C. Jiwan¹, Casey Appel¹, Rui Wang², Chwan-Li Shen^{2,3,4}, & Hui-Ying Luk¹

¹Department of Kinesiology and Sport Management, Texas Tech University, Lubbock, TX

²Department of Pathology, ³Center of Excellence for Integrative Health; ⁴Center of Excellence for Translational Neuroscience and Therapeutics, Texas Tech University Health Sciences Center, Lubbock, TX

TEXAS TECH UNIVERSITY
Department of Kinesiology
& Sport Management

TEXAS TECH UNIVERSITY
Department of Kinesiology
& Sport Management

ABSTRACT

With diabetes, skeletal muscle mitochondrial quality control (mitochondrial fusion, fission & macro-autophagy) is impaired. Geranylgeraniol (GG) is shown to have a protective effect on preventing mitochondrial damage and muscle health; however, the effect of GG on a diabetic model is not known. **PURPOSE:** To determine the effect of GG on mitochondrial quality control and muscle cross-sectional area (CSA) in diabetic rats. **METHODS:** Thirty-five Sprague-Dawley rats were divided into three diet groups: control diet (CON), high-fat diet with 35 mg/kg body weight of streptozotocin (HFD), and HFD with 800 mg/kg body weight of GG (GG). Due to the limited sample, a total of 21 (CON: n = 7; HFD: n = 7; GG: n = 7) rats' muscle samples were used for this report. The soleus muscles were harvested after 7-weeks of feeding and were analyzed for OPA1, MFN2, DRP1, pDRP, PINK1, Parkin, LC3A, and LC3B protein content using western blot analysis. Muscle CSAs were assessed using Image J. **RESULTS:** A significant ($p < 0.05$) condition effect was observed for MFN2, DRP1, LC3A, and LC3B protein contents and muscle CSA. For mitochondrial fusion, GG (0.21 ± 0.08) had lower MFN2 than CON (0.43 ± 0.04 ; $p = 0.007$) and HFD (0.65 ± 0.08 ; $p = 0.010$). For mitochondrial fission, GG (0.26 ± 0.07) had lower DRP1 than HFD (0.59 ± 0.07 ; $p = 0.019$). For macro-autophagy, GG (1.08 ± 0.28) had lower LC3A than CON (2.81 ± 0.55 ; $p = 0.028$) and HFD (3.99 ± 0.57 ; $p = 0.010$); whereas GG (0.63 ± 0.21) had lower LC3B than HFD (1.93 ± 0.24 ; $p = 0.012$). No significant differences were observed for OPA1, pDRP, PINK1, Parkin, and LC3B/A. For muscle size, CON ($10,092.88 \pm 104.67 \mu\text{m}^2$) had larger CSA than GG ($7284.69 \pm 70.91 \mu\text{m}^2$, $p = 0.001$) and HFD ($5615.59 \pm 59.97 \mu\text{m}^2$; $p = 0.001$), whereas GG ($7284.69 \pm 70.91 \mu\text{m}^2$) had larger CSA than HFD ($5615.59 \pm 59.97 \mu\text{m}^2$; $p = 0.001$). **CONCLUSION:** GG supplementation could prevent mitochondrial fragmentation (reduction in DRP1), thus, potentially resulting in a decreased demand for mitochondrial fusion (reduction in MFN2). In addition, a greater rate of autophagosome degradation than formation (reduction in LC3A and LC3B) was observed (indicative of an increase in macro-autophagy). Improvement in mitochondrial quality could potentially contribute to attenuating the reduction of muscle size in diabetic rats with GG supplementation.

INTRODUCTION

- Increased inflammation and oxidative stress can result in mitochondrial dysfunction, a potential pathogenic contributor to insulin resistance.
- Mitochondrial quality control (mitochondrial fission, fusion, & macro-autophagy) is a mechanism to maintain healthy mitochondria and prevent mitochondrial dysfunction.
- Individuals with Type 2 diabetes had increased fission (increased in DRP1), decreased fusion (decreased in MFN2), and reduced capacity to remove damaged mitochondria (decrease in PINK1, Parkin, and LC3B).
- Geranylgeraniol (GGOH) supplementation has been shown to reduce inflammatory markers and prevent mitochondrial damage in neuronal cells and preserve muscle cross-sectional area in the skeletal muscle.
- Improving mitochondrial quality is essential to improve metabolic regulation in diabetic populations; however, to date, the effect of GGOH on mitochondrial quality control and muscle cross-sectional area in a diabetic model is not known.

PURPOSE

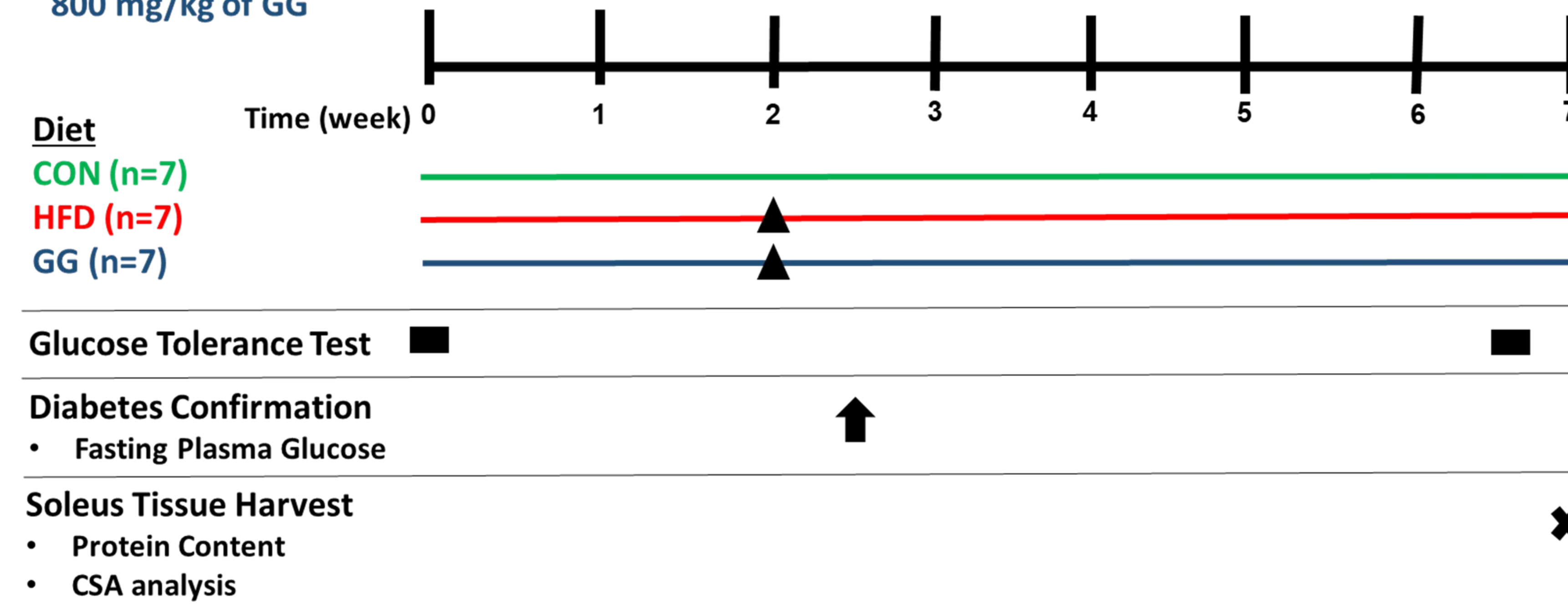
To determine the effect of GGOH on mitochondrial quality control and muscle cross-sectional area in diabetic rats.

METHODS

Brief Overview

Sprague-Dawley Rats (n=21)

- CON: Normal Diet
- High Fat Diet (HFD): HFD + 35 mm/kg of Streptozotocin (STZ)
- Geranylgeraniol (GG): HFD/STZ + 800 mg/kg of GG



Muscle Preparation and Protein Analyses

- Muscle samples were homogenized, agitated, and centrifuged.
- Protein content for mitochondrial fission (DRP1 & pDRP), fusion (MFN2 & OPA1), and macro-autophagy (mitophagy: PINK1 & Parkin; autophagy: LC3A & LC3B) were analyzed using western blot analysis. GAPDH was used as the loading control.
- Chemiluminescent substrate and the C-Digit imaging system were used to visualize the stained protein bands.
- Image Studio Digits Ver 4.0 was used for band densitometry.

Muscle Cross-Sectional Area (CSA) Analysis

- Muscle samples were sectioned at 10 μm followed by Hematoxylin and Eosin staining.
- Slides were visualized with a microscope and muscle CSA was analyzed using Image J.
- 100 muscle fibers from each rat were analyzed.

Data Analyses

- Data was analyzed using a one-way ANOVA.
- Bonferroni post hoc tests were used for pairwise comparisons.
- Statistical significance was set at $p \leq 0.05$.
- Data are reported as mean \pm SE.

RESULTS

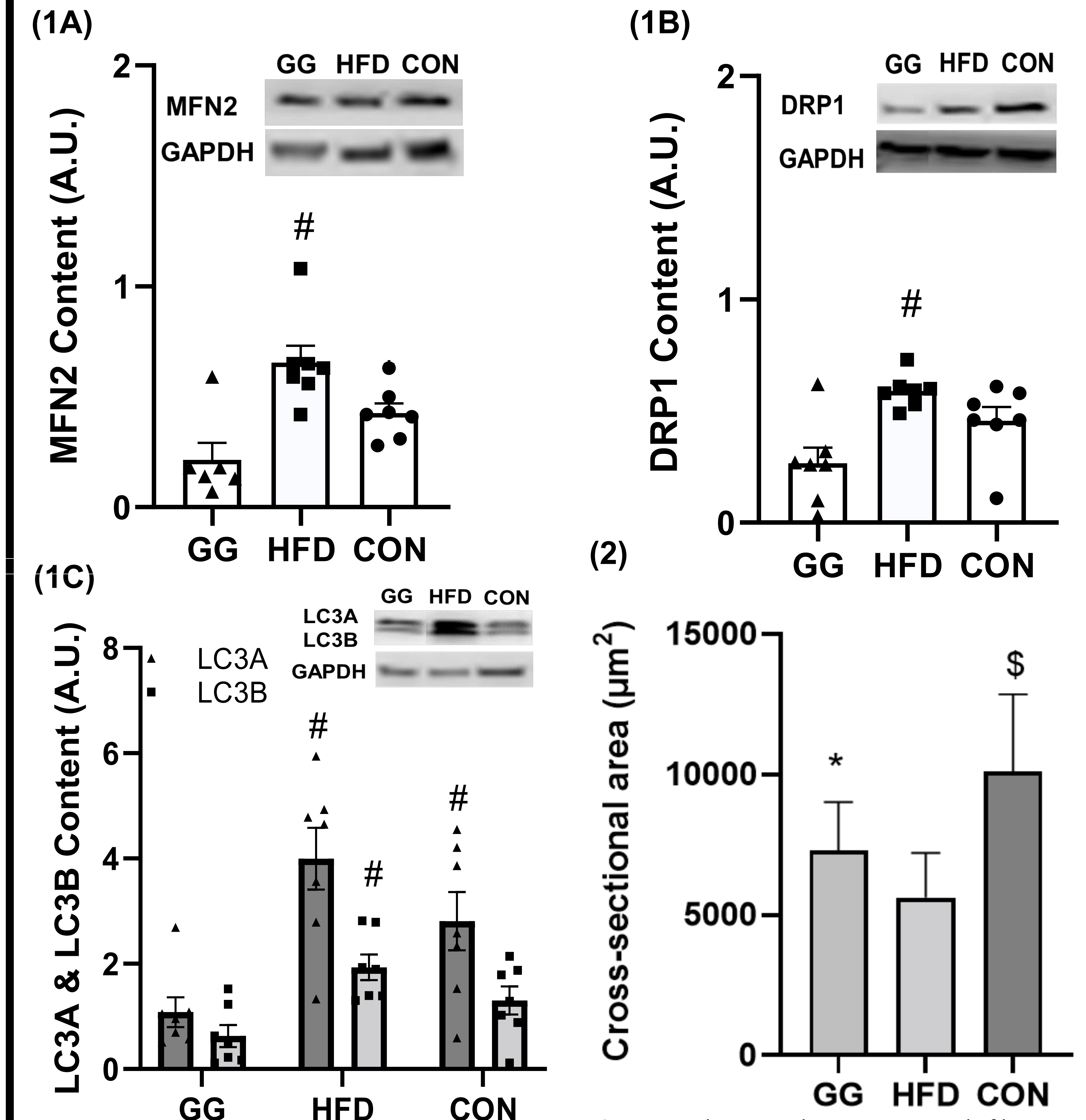


Figure 1. Protein content for (A) MFN2, (B) DRP1, (C) LC3A & B. A significant condition effect was observed for MFN2, DRP1, LC3A and LC3B. # $P < 0.05$ vs. GG

Figure 2. Soleus muscle CSA. 700 muscle fibers were counted for each group. A significant condition effect was observed for muscle CSA of the soleus. \$ $P < 0.05$ vs. GG and HFD. * $P < 0.05$ vs. HFD.

CONCLUSIONS

- Supplementing GGOH to diabetic rats was able to mitigate the CSA reduction observed in diabetic rats without GGOH.
- Additionally, a lower mitochondrial fragmentation (decrease in DRP1) and a greater autophagosome degradation (decreased in LC3A and LC3B) was observed in diabetic rats with GGOH when compared to diabetic rats without GGOH.

ACKNOWLEDGEMENTS

This project was supported by the by the Texas Tech University internal funding.

